Strategies to characterize Fungal lipase for application in Dairy industry and medicine

Sikander Ali*, Aneeqa Nasir, Sidra Sajjad, Fizza Siddique

Institute of Industrial Biotechnology, Government College University, Pakistan

*Corresponding author: Dr. Sikander Ali (Email: dr.sikanderali@gcu.edu.pk)

Abstract

Esters are synthesized by catalysis of lipolytic enzyme from fatty acids and glycerol. Lipases are extensively dispersed in the whole world, nevertheless merely lipolytic enzymes produced by microbes are significant commercially. The several uses of lipases includes hydrolysis of fats, taste enrichment in food treating, perseverance of racemic mixes and chemical evaluates etc. Lipases are used as analytical implements. Lipases are used in food industry as baking, brewing, dairy and cheese products formation, fruit juices, formation of syrup, starch production and especially in flavor production of cheese. They play an important part in curing, diagnosing, biochemical examination, and checking of several feared ailments. Lipases are used as indicator enzyme for medical purposes. Their increased or decreased level in the body show definite illness. They are used in drugs for diseases like Crohns disease, gastrointestinal disturbances cancer necrosis element and much more.

Keywords; Lipolytic enzyme, racemic mix, analytical, pancreatic lipase

Introduction

Lipolytic enzyme known as triacylglycerol acyl hydrolases. They harvest glycerol and free fatty acids by hydrolyzing fat and oil. (Singh & Mukhopadhyay, 2012). (Fig 1 shows conversion of triglyceride into glycerol and fatty acids). Lipases don't need any cofactor being a member of serine hydrolases. The Lipolytic enzyme involves in the catalysis of esters at the crossing point among anin solvable substrate part and liquid part at which enzyme leftovers liquefied (Ghosh *et al.*, 1996). Lipases take part in transformation reactions, for instance esterification, alcoholysis, inter-esterification, acidolysis, aminolysis, and transesterification (Savitha *et al.*, 2007). Numerous microbes for instance molds, bacteria yeasts and some protozoa are recognized to discharge the lipase enzymes for the assimilation of lipid constituents (Anbu *et al.*, 2011). Microorganisms, being universal in dissemination, are extremely fruitful at persisting in an extensive array of environmental situations because of their great elasticity and biological adaptability (Johri *et al.*, 1990).

1. Sources of lipases

Animals, plants and microorganisms are usually involved in the synthesis of lipases nevertheless lipases produced by microbes are considered to be technically significant later the lipases differentiated in their substrate specificity and properties (Ramakrishnan *et al.*, 2013). Lipases acquired by the animals primarily from pancreatic muscles of livestock. The drawbacks of utilizing lipases produced by animals involves the lipolytic enzyme of pig i.e., trypsin has unpleasant flavouring amino acids, also the hormones produced by these animals results in the dispensation of veggie (Vakhlu & Kour, 2006). Lipases produced by plants are not demoralized commercially owing to the harvest and the procedures. Consequently, lipases from are presently getting extra devotion due to their practical and financial rewards, where the organisms are nurtured in medium comprising suitable nutrients under organized environments (Treichel, 2010).

1.1. Lipase produced by bacteria

Most commonly the bacterial lipases are produced by *Bacillus pumilus, Bacillus subtilis, B. licheniformis, B. alcalophilus, Bacillus coagulans* and *Bacillus stearothermophilus*. Furthermore, Burkholderia multivorans, Pseudomonas aeruginosa, Staphylococcus caseolyticus and Burkholderia cepacia are too described equally as producers of bacterial lipase (Ertugrul *et al.*, 2007).



Fig: 1 Conversion of triglyceride into glycerol and fatty acids

1.2. Lipase produced by Fungi

The commercially significant lipase-manufacturing fungal genera includes Penicillium spp, Rhizopus spp, Geotrichum spp, Aspergillus spp, Rhizomucor spp and Mucor sp. Production of lipase by fungi fluctuates along with components in the growing medium, strain, growing conditions, temperature, availability of carbon ,pH and nitrogen sources (Cinhangir & Sarikaya, 2004). Thermophilic Mucor pusillus, Aspergillus terreus and Rhizopus homothallicus are the producers of extracellular thermostable lipase enzyme. Alkaliphilic, inducible, extracelluar and thermo-stable lipase enzyme is harvested by Mucor sp. There are limited knowledge about alkaliphilic and thermo-stable lipase production by molds.

2. Separation and Screening of lipolytic generating Fungi

The most suitable and consistent process of perceiving lipase activity in microbes involves using the surfactant Tween 80 in a compact form in order to observe the activity. This method has been discovered by Seirra in 1957. The appearance of misty regions is a sign of production of lipase by the microbes. Alterations to this process involves Tween surfactants amalgamation with Nile blue and copper salts. Similarly, selection of lipase manufacturers on agar medium normally performed by means of tributyrin as a substrate and formation of clear zones shows a signal of production of lipase (Cardenas *et al.*, 2001). Similarly, the Rhodamine process include usage of surplus water to regulate lipases synthesis by *Aspergillus ibericus* (Abrunhosa *et al.*, 2013). 32 different fungal species were isolated using

Rhodamine fluorescence-based assay. (Fig 2 indicates three plates comprising Tween-20, tributyrin and vegetable oil respectively and respective results on them).

3. Purification

Awareness about the refined lipase activities is the basis for biotechnological or supplementary applications. Several lipases from diverse microbes have been defined. Aspergillus niger produces lipases of molecular masses 31 and 19kDa (Hofel mann *etal.*, 1985), 30 kDa by *Rhizopus japonicas* (Suzuki *et al.*, 1986), 25kDa by *Aspergillus oryzae* (Toida *et al.*,1998) 35.5kDa by *Aspergillus niger* (Namboodiri and Chattopadhyaya, 2000) 49kDa by *Cunninghamella verticillata*. It looks that formation of lipolytic enzyme by different species of fungus effects in altered molecular weights, because of differences of quantity in the amino acid remains. (Fig 3 shows lipase purification by affinity chromatography). In general, old refining approaches are measured time-taking with poor harvests and the developments are stirring near purification in ionic liquids and aqueous two-phase extraction centered on lipase-lipase interface (Nagarajan, 2012).



Fig: 2 Media plates comprising substrates (a) Tween, (b) tributyrin, and (c) vegetable oil. Calcium crystals were seen in Tween agar plate. A clear zone was observed in Tributyrin plate, while development of luminescence was detected underneath Ultra Violet radiance.

4. Thermo-stability of lipase

Thermostable lipases have been purified from several microbes, comprising *B. coagulans* and *B. cereus P. fluorescens* (Kojima *et al.*, 1994); *B. stearo-thermophilus* (Kim *et al.*, 1998) *P. aeruginosa* and *Geotrichum* sp. and *Aeromonas sobria* (Lotrakul & Dharmsthiti, 1997), (Macedo *et al.*, 1997). *P. aeruginosa* lipase was knowingly steadied by Calcium ions and deactivated by Ethylene diamine tetraacetic acid. This deactivation overwhelmed by adding

Cacl₂, signifying the presence of an attachment site for calcium in *P. aeruginosa* lipase (El-Shafei & Rezkallah, 1997).



Fig: 3 Lipase purification by affinity chromatography

5. Immobilization of lipase

Together innate and immobilized lipases are accessible on commercial scale. They are being utilized in washing cleaners and several extra uses are not immobilized; nevertheless, a cumulative amount of uses of these enzymes in production and in biotransformation process claim a biocatalyst for efficacy of usage. Recyclibility of costly lipases recovers through immobilization. Also, constancy and activity of enzyme have been improved through immobilization. Numerous approaches have been castoff to immobilize lipases, comprising enchantment in gels that show polymerization (Telefoncu *et al.*, 1990), sorption or drizzle on water repellent resources (Wisdom *et al.*, 1984), sorption on anion interchange resins (Rizzi *et al.*, 1992), chemical bond formation (Shaw *et al.*, 1990) sol–gel enchantment (Krishnakant & Madamwar, 2001) and lipid cyst having micro-encapsulation. Accurel EP 100 porous polypropylene is used to immobilize *G. candidum* lipases A and B, precoated with ovalbumin to enhance constancy at raised temperatures. *R. miehei* lipase has been made immobilized through hydrophobic controlled pore glasses. Similarly various lipases have been made immobilized by sol–gel enchantment in silica gel (Reetz *et al.*, 1995).

6. Lipase gene cloning and sequencing

Lipase gene sequencing and cloning remains to entice devotion. Genes for lipase enzyme from numerous microbes and animals have been cloned. *Aci. calcoaceticus* BD413 DNA a

fundamental lipase gene transferred in *Escherichia coli* phage M13 (Kok *et al.*, 1995). Lipase from *Rhizopus oryzae* DSM 853 has cloned. Seemingly, *Rhizopus* sp. defined in the collected works effect from diverse proteolytic dispensation and has initiated the similar gene. Epitope mapping works by means of monoclonal antibodies are focused in contrary to human pancreatic lipase in addition many altered lipases propose that β -50 loop from C-terminal dominion might convoluted in collaboration with (HPL) along with a boundary (Bezzine *et al.*, 1998).L1, L2, and L3 are the extracellular lipolytic enzyme produced by *Aspergillus oryzae*. Out of these, L3 lipolytic gene was cloned (Toida *et al.*, 2000).

7. Application of lipase in dairy industry

The mandate of industrialized enzymes predominantly of microbial source is perpetually cumulative because of their uses in a diversity of manners. Reactions which are catalyzed by enzymes are striking and mostly chosen to chemical methods. Enzymes are used in widespread variation of arenas including food industry as baking, brewing, dairy and cheese products formation, fruit juices and also the formation of syrup and starch production. It can be perplexing to find a proper enzyme because one enzyme can be utilized in several means for example role of lipases, by interesterification, altering of fats takes place in which palm oil with stearic acid used in chocolate confectionery by fat hydrolysis for the production of aromatic flavors and for cheese maturation.

Lipolytic enzymes are included in the class of hydrolases and cause the breakdown of acylglyceride. The lipases catalyze wide-ranging of reactions, counting hydrolysis, alcholysis, aminolysis, inter-etherification, acidolysis, esterification and. Role of lipases as taste improvement negotiators in cheese formation, and in fat and cooking oil is important (Vakhlu, 2006). Many exogenous enzymes and indigenous milk enzymes are tangled in the production and seasoning of cheese, and to hydrolyze milk fat for the development of flavors, this speed up cheese maturing and to enhance the flavourings for cheese for example cheese equivalents (Woo & Lindsay, 1984). Lipase which was artificially produced in *A. oryzae* was used as processing support in the baking industry (Greenough *et al.*, 1996).

8. Role of lipase in cheese production

According to FAO, "cheese" is defined a renewed merchandise or a seasoned artifact which is formed by portly milk, condensing milk, cream, sped-read milk or moderately sped-read milk, or a limited or full blend of produce, monitored by settling whey (Syoten, 1963). There are two types of cheese which are most common; Production of cheese through thickening of milk. Milk coagulation in cheese takes place by enzymes, removed from the stomach of fresh calves or from other bases (rennet) is best.Mainly texture and flavor of cheeses depends on the fat content of the milk. Cheeses are of four types on the basis of fat concentration. Little fat content includes (<25 %), Moderate fat content includes (25-45%), Complete fat content includes (45-60%), Extraordinary fat content includes (>60%). Diversities in mould seasoned cheeses are especially Camembert, Brie, Roquefort and Stilton having widespread lipolysis and distinct flavor and aroma.

9. Flavor development of Cheese

Primary and secondary biochemical ways including proteolysis, lipolysis, breakdown of residual lactose and of lactate and citrate and breakdown of fatty acids and amino acids, many compounds are manufactured which have a role in cheese flavorings (Marilley & Casey, 2004). Cheese lipases and enzymes essential in curdled milk of unweaned calf are elaborate in lipolysis in cheeses like Greek Feta, Italian Pecorino cheeses and Provolone. Subordinate responses from breakdown of fatty acids and amino acids are involved for the creation of volatile savor combinations. Growing of cheese matures their distinguishing savor through continuing itemization of proteins, lipids and carbohydrates. In best ripened cheese assortments, enzymes from cheese microbes especially lactic acid bacteria are elaborate in flavour improvement. Currently, some distinctive starter cultures recognized as adjunct cultures be situated together with the key initiator cultures to hasten flavor expansion and other fermented foods and also rush cheese growing, permit extensive total investments to be saved by the cheese industry. Role of microbial lipases is shown in table: 1.

10. Enzyme modified cheese flavorings

The EMC production is the application of certain enzymes for the development of strong cheese flavor quickly from dairy substrates under optimal environments. In the late 1960s the knowledge in which new curd and solution of NaCl were used to create a mixture having ~ 40% total artifacts (Kilcawley & Fox, 1998). Preservers and lipolytic enzymes added in the mixture and conditions were also carefully controlled to give the maturation time to mature flavour. Marketable manufacture of EMC involves the use of definite enzymes and/or microorganisms which are involved in the production of developed or undeveloped cheese and heating system is used to stop this practice and then end product was standardized on the way to a favorite aroma strength and composition. Production of EMC is shown in figure:4.

Table: 1 Role of microbial lipases in Food Industry

Industry	Effect	Product
Bakery	Flavor and shelf life improvement	Bakery
Beverages	Better-quality aroma	Beverages
Dairy	Milk fat hydrolysis	Flavor compounds
	Cheese maturing	Coagulated milk
	Butter fat alteration	Fat
Fats and oils	Trans-esterification Water breakdown	Cocoabeans,Butter,Cookingoil,Fattyacids,Glycerol,monoanddiglycerides
Food dressing	Quality perfection	Mayonnaise, vinaigrettes and floggings
Meat and fish	Flavor increase and plump elimination	Kernel and fish artifacts
Health food	Transesterification	Health food



Fig: 4 Production of cheese flavor (EMC) and butter flavor (EMB)

EMC may be produced by two ways. One way is involved only single stage in which fat and protein breakdown happen at the same time and in other way different aroma constituents are produced independently and finally blend them. The real handling circumstances be governed by mainly the enzymes which are used. About 12 - 72 hours, $30 - 45^{\circ}$ C temperature and pH 5-7 is required for enzyme modified cheese flavorings development. (Kilcawley & Fox, 1998). In order to improve its storage stability, or flavour the pH of EMC may be altered while bulking agents or carriers may be utilized to assist sprig drying and yield enhancement as well as desired aroma. Creams or precipitates which are obtained as end harvests have high focusses of peptides, free amino acids, and other artifacts which can be broken down, usually accompanying with aroma of cheeses. Lipolysis is involved mainly in EMC flavour because of the development of extraordinary ranks of small series volatile fatty acids. Many of the EMC flavors are produced for Swiss type, Gouda type, Parmesan type, Mozzarella type and for Blue type cheeses.

11. Enhancement of cheese ripening

Cheese Ripening is a measured and luxurious method and its maturing time is four weeks to twenty eight months which is dependent on which type of cheese is used. In order to fasten the cheese maturation following methods are mostly used; Maturation at high temperatures, Lipolytic enzyme accumulation (e.g. pregastric esterase, the main lipolytic mediator for Italian cheeses), Enzyme-improved cheese and Cheese mixtures, Physically and Chemically altered microbial cells, Inherently improved starter cultures, Use of subordinate culture (Roginski *et al.*, 2003).

12. Types of Cheeses

12.1. Swiss-type

Initially they have developed in Emmental in Switzerland. Emmental is possibly the best cheese and known as 'Swiss cheese' (Bachmann *et al.*, 2003). They exhibit distinctive nutlike, sweet flavour. The water-soluble portion comprehending amino acids proline and valine are responsible for sweet flavor (Hintz *et al.*, 1956). In this cheese lactic acid, heat tolerant bacteria e.g *Str. thermophilus* and L. helveticus and propionic acid bacteria (PAB), commonly *Propionibacterium freudenreichii* spp. shermanii are used as first course cultures. Propionic acid bacteria is used to develop the particular aroma of this cheese that

differentiates it from other cheeses. N-caproic acids, isovaleric, n-butyric, propionic and acetic acids are responsible for distinct flavors of this cheese particularly (Ha & Lindsay, 1990, Ji *et al.*, 2004).

12.2. Italian cheese

In Italian cheese production fatty acids blends are required for their particular flavor (Ganesan & Weimer, 2004). Before the starter culture, Lipolytic enzyme from *Mucor miehei* is added in milk so precise lipolysis occur to develop different flavors as faint and fatty taste of Asiago Mozzarella, pungent taste of provolone and severe piccante of Romano (Italian cheeses).

12.3. Cheddar

It is a long-matured because nine months to two years for the production of its characteristic flavor are necessary. Fom milk fat degradation fatty acids for example caproic, capric and butyric acids are produced which are involved in its flavor production. Acidity of Cheddar cheese is due to the production of acetic and propionic acids as well as lactic acid. In Cheddar and Provolone cheeses development, gastric lipase of calf and pregastric lipase of goat together because their superiority is greater in combination as compared to pregastric lipase used unaccompanied (Lai *et al.*,1997).

12.4. Mold or Smear seasoned cheeses

In these cheeses lipids breakdown of lipids is due to the enzymes from the mould and may be from other subordinate cultures (Collins *et al.*, 2003). The distinctive spicy savor of Blue cheese is due to the production of methyl ketones and fatty acids having short chains and the *Penicillium roqueforti* lipase is involved in its lipolysis with a small influence of indigenous milk lipase.

13. Role of lipases in medicine

The chief enzyme involved in the breakdown of fats is the lipolytic enzyme and its scarcity cause terrible significances to fitness. The lipolytic enzyme is an initiator for cancer necrosis element and has been used for the cure of malevolent growths (Stolzmann & Mikolajczak, 2001). Other beneficial uses of lipolytic enzyme with additional constituents known to cure gastrointestinal disturbances, dyspepsias, cutaneous appearances of gastric antipathies and many more. Retarding the working of microbial lipase in order to control infection or to regulate fatness, promoting the action of lipolytic enzyme to lessen procoagulant state or

hyperlipidemia, or utilizing lipid hydrolyzing enzyme as complement to manage Crohn's disease(as shown in fig. 5) also known as cystic fibrosis, completely associated with human fitness. Celiac syndrome; an ailment in which gluten harms the duodenal tractso strict diet free of gluten is taken by patients. Pancreatic enzymes is used as a medication for celiac disease (Hegnhoj *et al.*, 1990). Lipases are being utilized as analytical implements as well as increase in its concentrations show definite illness. These are significant goals for drugs and indicator enzymes for medical purposes. Blood serum having specified amount of lipid hydrolyzing enzymes are recycled by way of an analytical instrument to detect injury of pancreas and severe pancreatitis (Shepherd, 1993). Lipase level is too significant in making the judgment of heart sicknesses (Nikkila *et al.*, 1977). Lipolytic enzyme might restrained at O_2 /pH cathodes through glucose oxidase, play a role of fat biosensors as well as in blood cholesterol determinations (Mukherjee, 2000).

14. Lipase in pancreas

The lipase enzyme present in pancreas is exocrine enzyme (EC 3.1.1.3) of juice in pancreas and play a significant role in the breakdown of fats in duodenal lumen. Water breakdown of the triacylglycerols involves pancreatic and gastric lipolytic enzyme significant aimed at their immersion through enterocytes, so that fat is absorbed in the body (Esposito, 1973). Its substrate is not a single molecule but accumulated lipids that are composed of ester particles and interface with a liquid medium (Lagocki *et al.*, 1973) The lipase in pancreas needs colipase (a protein in pancreas)as cofactor for the commotion of enzyme. Pancreatic lipase that is colipase dependent enables the endorsement of fatty acids while lipolytic enzyme motivated bile salts aids in acceptance of permitted lipid molecules through the lumen of intestine (Brockman, 2000).



Fig:5 This figure shows the shape of ileum in normal and in diseased(crohns disease) condition.

15. Use of digestive lipase preparations

Majority people suffering from cystic fibrosis disease need enzyme in the pancreas as complements daily comprising lipolytic, carbohydrate degrading and protein degrading enzymes, to dispense with indications of exocrine pancreatic insufficiency. The use of complementations of pancreatic enzyme is beneficial for treatment of people suffering from celiac disease (a disorder in which duodenal region is infected as well as cause the less absorbtion of the nutrient). Crohn's disease, associated with lack of enzymes in the pancreas. Lipolytic enzymes as well as additional enzymes of pancreas supplementations are present in the form of medicine (Hegnhoj, 1990).

16. Inhibition of lipase activity

16.1. Defiant-microbial accomplishment of lipase

The extracellular commotion of lipase allows infectious organisms to cast-off lipids over skin and mucosal coatings and grow on these surfaces i.e the pathogen of humans *Candida albicans*, contaminates the skin and mucosa along with unadorned general contamination (Stehr et al., 2000). The inhibitors of the lipase found in microbes are capable to regulate toxicities produced by microorganisms that secrete lipolytic enzyme. Usual artifacts for example, related alkaloids sanguinarine and berberine known to have anti-lipolytic commotion compared to *C. rugosa* lipolytic enzyme (Hube *et al.*, 2000).

16.2. Anti-obesity agents

Obesity has a dangerous aspect on syndromes for example, hyperlipidemia, hypertension cardiovascular disease and diabetes mellitus. As hydrolysis of dietary triacylglycerol is important for successive absorption by enterocytes, anti-lipase agents work by decreasing or hindering the availability of dietary fat calories by stopping absorption of fats, and thus mimic the influence of reduced food intake (Bray, 2000). First and foremost effective inhibitors of lipolytic enzyme in pancreas include lipstatin, it was sequestered by *Streptomyces toxytricini*, its beta lactone assembly inhibit pancreatic lipase (Weibel *et al.*, 1987). Tetrahydrolipstatin is a synthetic compound and was recycled for the cure of fatness (Haputman *et al.*, 1992). Extract of herb name Nomame herba was lately testified to hinder porcine pancreatic (Toplak & Marhardt, 1998). Enterostatin cause reduction of weight, it is

formed by pancreatic procolipase and is produced in gastrointestinal lumen by high fat diet. It is secreted less in obese persons. Its anorectic effect on CNS and PNS of brain results in numerous mechanisms of breakdown (reduction in insulin, increase in corticosteroid and adrenal excretion as well as improved compassionate initiative) to brunette adipose muscles (Albertsson & Enterostatin , 1997).

17. Stimulation of lipase activity

Hyperlipidemia for instance in Fredrickson Type III, IV or IIb has a menace element for precipitate atherosclerosis Hypertriglyceridemia might occur through decrease in elimination of mingling triglyceride because of decrease in commotion of lipolytic enzyme of lipoprotein, or through the higher level of hepatic secretion of triglyceride-rich VLDL. Activation of lipoprotein lipase activity is done to lessen the triglyceride levels and hydrolysis occur for catalysis (Shepherd, 1998).

17.1. Fibrate drugs

They are deliberated to be supreme cooperative in decreasing triglyceride intensities in serum and their mode of action is improvement of lipoprotein lipase activity. They also cause: in the muscle there is high approval and corrosion of fatty acids. Reduction in production along with discharge of VLDL through liver. Enlarged production of Apolipoprotein A1. Lowering the expression of apolipoprotein CIII gene (Auwerx *et al.*, 1996).

17.2. Simvastatin (statin drug)

Simvastatin is an inhibitor of 3-hydroxy-3-methylglutaryl-coenzyme. It is related in the treatment of hyperlipidemia especially hypertriglyceridemia (Stolzmann & Mikolajczak, 2001).

17.3. Heparin

Heparin lipolytic activity is systematized through the discharge of hepatic lipase and lipoprotein (Olivecrona &Bengtsson, 1989). Heparin function as lipase and also facilitates the discharge of chockfull length tissue factor pathway inhibitor into rotation. Post-heparin plasma development through antibody alongside the C-terminal section of Tissue Factor Pathway Inhibitor introverted the lipoprotein lipase commotion (Mukherjee & Kakkar, 1999).

18. Cancer related cachexia

Cancer linked cachexia is complex multiorgan disorder frequently related with various forms of cancer. Cachexia is the reason of death of 15-20 % patients having cancer. The level of cachexia changes depending on the type of the tumor site and stage patients that have CAC show anorexia and decrease of body weight leading to physical weakening. Patients experience loss of skinny tissues and adipose muscle (as shown in fig: 6). Cancer changes metabolism of lipid into catabolism therefore decreasing the level of fat mass lipid metabolism is highly changed and cause lipotoxic effect in tissues including skeletal muscle. Lipoprotein lipolytic enzyme belongs to family of triglyceride hydrolase as well as is attached to the endothelial cell surface of the white adipose tissue ,heart and skeletal muscles. It hydrolyses TAG.Adipose triglyceride lipase catalyses the conversin of tag into diacylglycerol and a free fatty acid.thus causing increased lipolysis that was previously considered as cell death (Das and Gerald ,2013).



Fig: 6 Mechanism of lipase action in cancer related Cachexia

19. Conclusion

Fungi are able to produce lipases on behalf of their existence inside a varied kind of substrates which are used in many applications. Lipases are striking for their uses in areas applicable to medication and dairy industry. The major benefits of lipolytic enzymes produced by fungus involves that they are effortlessly extracted in pure form i.e., extracellular nature. Lipase-centered handling has a hopeful future; though, the proportion of development is slow. Causes for restrictions comprise a moderately great cost of lipolytic enzyme and lack of enzymes through optimum series of catalytic activities essential for many applications.

References

- Abrunhosa, L.,Oliveira, F., Dantas, D., Goncalves, C., & Belo, I. (2013). Lipase production by Aspergillus ibericus using olive mill wastewater. *Bioprocess and Biosystem Engineering*, 36, 285–291. Retrieved April 4, 2017.
- Albertsson, C.E., & York, D. (1997). Enterostatin: a peptide regulating fat intake. *Obes. Res*, 5, 360–372. Retrieved March 27, 2017.
- Anbu,P.,Noh,M., Kim,D.,Seo,J., Hur,B.,& Min,K.H.(2011). Screening and optimization of extracellular lipases by Acinetobacter species isolated from oil-contaminated soil in South Korea. *African Journal of Biotechnology*, 10 (20), 4147–4156. Retrieved March 29, 2017.
- Anbu,P., Gopinath,S.C.B., Hilda,A.,Priya,T.L., & Annadurai,G. (2007). Optimization of extracellular keratinase production by poultry farm isolate Scopulariopsis brevicaulis.
 Bioresource Technology, 98(6), 1298–1303. Retrieved March 24, 2017.
- Auwerx, J.,Schoonjans,K., Fruchart, J.C., & Staels, B. (1996). Transcriptional control of triglyceride metabolism: fibrates and fatty acids change the expression of LPL and apo C-III genes by activating the nuclear receptor PPAR, Atheroscler. *Suppl*, 124, 29– 37. Retrieved March 24, 2017.
- Bachmann, H.P., Butikofer, U., & Isolini, D. (2003). Swiss-type cheese United Kingdom: Academic press.
- Bezzine,S., Carriere, F., Decaro,J., Verger, R., and Decaro,A. (1998). Human pancreatic lipase: an exposed hydrophobic loop from the C-terminal domain may contribute to interfacial binding. *Biochemistry*, 25,1846–1855. Retrieved April 6, 2017.
- Bray,G.A. (2000).A concise review on the therapeutics of obesity. *Nutrition*, 16,953-960.Retrieved March 25, 2017.
- Brockman, H.L. (2000). Kinetic behaviour of the pancreatic lipase– colipase–lipid system. *Biochimie*, 82, 987–995. Retrieved April 4, 2017.
- Cardenas, J., Alvarez, E., Alvarez, C.M.S., Sanchez, J.M., Valmaseda, M., Elson, S.W., & Sinisterra, J.V.(2001). Screening and catalytic activity in organic synthesis of novel fungal and yeast lipases. *J Mol Catal B: Enzym*, 14,111–23. Retrieved March 25, 2017.

- Cinhangir, N., & Sarikaya., E. (2004). Investigation of lipase production by new isolated Aspergillus sp. *World journal of Mocrobiology and Biotechnology*, 20, 193-197. Retrieved April 5, 2017.
- Collins, Y. F., Mc-Sweeney, P.L., & Wilkinson, M.G. (2003). Evidence of a relationship between autolysis of starter bacteria and lipolysis in Cheddar cheese during ripening. *J Dairy Res*, 70,105–113. Retrieved April 11, 2017.
- Das, S.K., & Gerald,H.(2013).Role of triglyceride lipase in cancer associated cachexia.*Trends in moleculare medicine*,19 (5),292-301. Retrieved April 9, 2011.
- El-Shafei,H.A., & Rezkallah, L.A.(1997). Production, purification and characterization of Bacillus lipase. *Microbiol Res*, 52,199–208. Retrieved March 21, 2017.
- Ertugrul, S., Donmez, G., & Takaç, S. (2007). Isolation of lipase producing Bacillus sp. from olive mill wastewater and improving its enzyme activity. *Journal of Hazardous Materials*, 149, 720–724. Retrieved March 24, 2017.
- Esposito,S., Semeriva, M., & Desnuelle, P.(1973). Effect of surface pressure on the hydrolysis of ester monolayers by pancreatic lipase. *Biochim Biophys Acta*, 302, 293–304. Retrieved March 27, 2017.
- Ganesan, B., & Weimer, B.C. (2004). Role of aminotransferase. In: Production of branchedchain fatty acids by Lactococcus lactis subsp. lactis. *Appl Environ Microbiol*, 70,638-641. Retrieved April 3, 2017.
- Ghosh, P.K., Saxena, R.K., Gupta, R., Yadav, R.P., & Davidson, S. (1996). Microbial lipases: production and applications. *Scienceprogress*, 79, 119–157. Retrieved April 13, 2017.
- Greenough, R. J., Perry, C.J., & Stavnsbjerg, M. (1996). Safety evaluation of a lipase expressed in Aspergillusoryzae, *Food ChemToxicol*, 34, 161-166. Retrieved March 21, 2017,
- Guo,Q., Wang,P.R., Milot,D.P., Ippolito, M.C., Hernandez, M., Burton, C.A., Wright, S.D., & Chao,Y. (2001). Regulation of lipid metabolism and gene expression by fenofibrate in hamsters. *Biochim Biophys Acta*, 1533, 220–232. Retrieved April 2, 2017.
- Ha, J. K., & Lindsay, R.C. (1990). Method for the quantitative analysis of volatile free and total branched-chain fatty acids in cheese and milk fat. *J Dairy Sci*, 73, 1988–1999. Retrieved April 7,2017.

HRD🗐

- Haputman, J.B., Jeunet, F.S., & Hartmann, D.(1992). Initial studies in human with the novel gastrointestinal lipase inhibitor Ro 18-0647 (tetrahydrolipstatin). Am. J. Clin. Nutr, 55, 3095-3135. Retrieved April 16, 2017.
- Hegnhoj,J., Hansen,C.P., Rannem,T., Sobrik,H., Anderson,L.B., & Anderson,J.R.(1990). Pancreatic function in Crohn's disease. *Gut*, 37,1076–1079. Retrieved March 27, 2017.
- Hofelmann,M., Hartmann,J., Zink,A., & Schreier,P.(1985). Isolation, purification and characterization of lipase isozymes from a technical Aspergillus niger enzyme. *Journal of Food Science*, 50, 1721–1726. Retrieved March 18, 2017.
- Hube,B., Stehr,F., Bossenz,M., Manzur,A., Kretschmar, M., & Schafer, W.(2000). Secreted lipases of Candida albicans: cloning, characterization and expression analysis of a new gene family with at least 10 members. *Arch. Microbiol*, 174, 362–374. Retrieved April 7, 2017.
- Hintz, P.C., Slater, W.L., & Harper, W.J. (1956). A survey of various free amino acids and fatty acids in domestic Swiss cheese. J. Dairy Sci, 39, 235–244. Retrieved March 17, 2017.
- Ji, T. V., Alvarez, B., & Harper, W.J. (2004). Influence of starter culture ratios and warm room treatment on free fatty acid and amino acid in Swiss cheese. J Dairy Sci, 87, 82-92. Retrieved March 21, 2017.
- Johri, B.N., Alurralde, J.D., & Klein,J. (1990). Lipase production by free and immobilized protoplasts of Sporotrichum (Chrysosporium)thermophile apinis. *Applied Microbiology and Biotechnology*, 33(4), 367–371. Retrieved April 14, 2017.
- Kashyap,M.(1984). The effect of gemfibrozil on plasma lipids and lipoproteins in man. *Vasc Med*, 2, 16–21. Retrieved April 17, 2017.
- Kim, H.K., Park, S.Y., Lee, J.K., & Oh, T.K. (1998). Gene cloning and characterization of thermostable lipase from Bacillus stearothermophilus L1. *Biosci Biotechnol Biochem*, 62,66–71. Retrieved March 26, 2017.
- Kilcawley, K. M. G., & Fox, P.F. (1998). Enzymemodified cheese. *Int Dairy J*, 8,1-10. Retrieved March 28, 2017.

- Kojima,Y., Yokoe, M., & Mase,T. (1994). Purification and characterization of alkaline lipase from Pseudomonas fluorescens AK102. *Biosci Biotechnol Biochem*,58, 1564–1568. Retrieved April 13, 2017.
- Kok, R.G., Thor,J.J.V., Roodzant, I.M.N., Brouwer, M.B.W., Egmond, M.R., Nudel,C.B., Vosman,B., & Hellingwer,K.J.(1995). Characterization of the extracellular lipase lipA of Acinetobacter colcoaceticus BD413 and sequence analysis of the cloned structural gene. *Mol Microbiol*, 15,803–18. Retrieved March 24, 2017.
- Krishnakant, S., & Madamwar, D. (2001). Ester synthesis by lipase immobilized on silica and microemulsion based organogels (MBGs). *Process Biochem*, 36, 607–11. Retrieved April 17, 2017.
- Lai, D. T., Mackenzie, A.D., O' Conno,C.J., & Turener,K.W. (1997). Hydrolysis characteristics of bovine milk fat and monoacid triglycerides mediated by pregastric lipase from goats and kids. *J Dairy Sci*, 80, 2249–2257. Retrieved April 12, 2017.
- Lagocki,J., Law J., & Kezdy,F.(1973). The kinetic study of enzyme action on substrate monolayers. Pancreatic lipase reactions. J. Biol Chem, 248, 580–587. Retrieved March 29, 2017.
- Lotrakul, P., & Dharmsthiti, S. (1997). Lipase production by Aeromonas sobria LP004 in a medium containing whey and soybean meal. *World J Microbiol Biotechnol*,13, 163– 166. Retrieved April 11, 2017.
- Macedo, G.A., Park,Y.K., & Pastor,G.M. (1997). Partial purification and characterization of an extracellular lipase from a newly isolated strain of Geotrichum sp. *Rev Microbiol*, 28,90–5. Retrieved April 19, 2017.
- Marilley, L., & Casey, M.G. (2004). Flavours of cheese products: Metabolic pathways, analytical tools and identification of producing strains. *Int J Food Microbiol*, 90,139-159. Retrieved April 22, 2017.
- Mukherjee, M., & Kakkar, V.V. (1999). Enhancement of lipoprotein lipase activity by tissue factor pathway inhibitor. *Thromb Haemost*, 82, 1648–1651. Retrieved March 31, 2017.
- Mukherjee, M., Ranlall, N., Patel, G., Rutlin, A., Jehanli, A., & Kakkar, V.V. (2000). Anticoagulant versus amidolytic activity of tissue factor pathway inhibitor in coronary artery disease. *Blood Coagul Fibrinol*, 11, 285–291. Retrieved March 24, 2017.

- Namboodiri, V.M. H., & Chattopadhyaya, R. (2000). Purification and biochemical characterization of an ovel thermostable lipase from Aspergillus niger. *Lipids*, 35 (5), 495–502. Retrieved April 19, 2017.
- Ngarajan, S. (2012). New tools for exploring, old-friend microbial lipases. *Applied Biochemistry and Biotechnology*, 168, 1163–1196. Retrieved April 13, 2017.
- Nikkila,E.A., Huttunen,J.K., & Ehnholm,C.(1977). Effect of clofibrate on post-heparin plasma triglyceride lipase activities in patients with hypertriglyceridemia. *Metabolism*, 26, 179–186. Retrieved March 21, 2017.
- Olivecrona, T., & Olivecrona, G.B. (1989). Heparin and lipases, in: D.A. Lane, U. Lindahl (Eds.), Heparin, Chemical and Biological Properties Clinical Applications. *Edward Arnold London*, 335-361. Retrieved April, 15 2017.
- Ramakrishnan, V., Goveas, L.C., Narayan,B., & Halami,P.M.(2013). Comparison of lipase production by Enterococcus faecium MTCC, 5695 and Pediococcus acidilactici MTCC, 11361 using fish waste as substrate: optimization of culture conditions by response surface methodology. *ISRN Biotechnology*, 980(62), 76-89. Retrieved April 22, 2017.
- Reetz, M.T., Zonta,A., & J.Simpelkamp.(1996). Efficient immobilization of lipases by entrapment in hydrophobic sol-gel materials. *Biotechnol Bioeng*, 49,527–34. Retrieved April 3, 2017.
- Rizzi, M., Stylos, P., Rick, A., & Reuss, M.(1992). A kinetic study of immobilized lipase catalyzing the synthesis of isoamyl acetate by transesterification in n-hexane. *Enzyme Microb Technol*, 14,709–13. Retrieved March 30, 2017
- Roginski,H., Fuquay,J.W., Fox,P.F.(2003).Encyclopedia of dairy science. *Academic Press,UK*, 252-444. Retrieved March 25, 2017.
- Savitha, j., Srividya,S., & Jagatetal,R. (2007). Identificationofpotential fungal strain(s) for the production of inducible, extracellular and alkalophilic lipase. *African Journal of Biotechnology*, 6(5), 564–568. Retrieved April 23, 2017.
- Shaw, J.F., Chang, R.C., & Wang,H.J.(1990). Lipolytic activities of a lipase immobilized on six selected supporting materials. *Biotechnol Bioeng*, 35,132–137. Retrieved March 27, 2017
- Shepherd, J.1993. Mechanism of action of fibrates. Postgrad. Med. J., 69:34-41.

- Singhand ,A.K., & Mukhopadhyay,M.(2012). Overview of fungal lipase: a review Applied Biochemistry and Biotechnology. *Science*, 106 (2),486–520. Retrieved April 11, 2017.
- Stehr,F., Felk, A., Kretschmar,M., Schaller,M., Schafer,W., & Hube,B.(2000). Extracellular hydrolytic enzymes and their relevance during Candida albicans infection. *Mycoses*, 43, 17–21. Retrieved April 14, 2017.
- Stolzmann,M.P., & Mikolajczak,A.P.(2001). The influence of simvastatin on lipase and cholesterol esterase activity in the serum of men with coronary heart disease. *Pharmacol Res*, 43,359–362. Retrieved March 24, 2017.
- Suzuki, M., Yamamoto, H., & Mizugaki, M. (1986). Purification and general properties of a metal-insensitive lipase from Rhizopus japonicus NR 400. *Journal of Biochemistry*,100(5), 1207-1213. Retrieved March 17, 2017.
- Syoten, A. (1963). Dairy product manufacture. I. Editing Committee for Dairy Technology Series Books, 1st edn, Oct 30, Tokyo Sztajer H, Bryjak M 1989, Capillar membranes for purification. of Pseudomonas fluorescens lipase. *Bioprocess Eng*, 4,257–259. Retrieved April 24, 2017.
- Telefoncu, A., Dinekaya, E., & Vorlop,K.D.(1990). Preparation and characterization of pancreatic lipase immobilized on Eudragit matrix. *Appl Biochem Biotechnol*, 26,311– 317. Retrieved March 17, 2017.
- Thompson,G.R.(1990). Primary hyperlipidemia. *Br Med Bull*, 46, 986–1004. Retrieved April 24, 2017.
- Toida,Y., Arikawa,K., Kondou,M., Fukuzawa, M., & Sekiguchi,J.(1998). Purification and characterization of Triacyl glycerol lipase from Aspergillus oryzae. *Bioscience Biotechnology and Biochemistry*, 62(4), 759–763. Retrieved April 6, 2017.
- Toida, J., Fukuzawa,M., Kobayashi,G., & Sekiguchi,J.(2000).Cloning and sequencing of triacylglycerol lipase gene of Aspergillus oryzae and its expression in Escherichia coli. *FEMS Microbiol Lett*, 189, 159–64. Retrieved April 4, 2017.
- Toplak,H., & Marhardt,K.(1998). Reduction of obesity and improvement in metabolic parameters by inhibition of intestinal lipases: current results with Orlistat. *Acta Med Aust*, 25, 142–145. Retrieved April 21, 2017.

- Treichel, H., de Oliveira, D., Mazutti, M.A., Di Luccio, M., & Oliveira, J.V. (2010). A review on microbial lipases production. *Food and Bioprocess Technology*, 3(2),182-196. Retrieved Maech 22, 2017.
- Vakhlu, J and A. Kour. 2006. Yeast lipases: enzyme purification, biochemical properties and gene cloning. *Electronic Journal of Biotechnology.*, 9(1): 69–81.
- Vakhlu, J.(2006). Yeast lipases: Enzyme purification, biochemical properties and gene cloning. *Biotechnol Ind*, 9, 53-59. Retrieved March 25, 2017.
- Weibel,E.K., Hadvary,P., Hochuli,E., Kupfer,E., Lengsfeld,H.(1987). Lipstatin, an inhibitor of pancreatic lipase, produced by Streptomyces toxytricini. I. Producing organism, fermentation, isolation and biological activity. *Antibiot*, 40 ,1081–1085. Retrieved April 18, 2017.
- Wisdom, R.A., Dunnill,P., Lilly,M.D., & Macrae,R.A.(1984). Enzyme esterification of fats: factors influencing the choice of support for immobilized lipase. *Enzyme Microb Technol*, 6, 443–446. Retrieved April 16, 2017.
- Woo, A.H., & Lindsay, R.C.(1984). Concentrations of major free fatty acids and flavor development in Italian cheese varieties. *J Dairy Sci*, 67, 960-968. Retrieved April 7, 2017.